

Infrageneric relationships within *Pogostemon* Desf. (Labiatae)

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Cladistic and phenetic analyses of the morphology of 79 species of *Pogostemon* Desf. sensu lato have been undertaken. The cladistic analysis included 135 character state differences encoded as 41 binary or multistate characters. The phenetic analysis included 52 metric variables and 54 binary variables. There is a very good correspondence between the results of cladistic and phenetic analyses based on different data sets. The existence of several clearly marked clades/clusters has been demonstrated. Some groups/clades are similar to those previously taxonomically recognized but the relationships and membership of infra-generic groups have been clarified by these analyses, validating a revised infra-generic classification which has been proposed.

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ADDITIONAL KEY WORDS:—cladistics – Dysophylla – Eusteralis – Lamiaceae – morphology – phenetics.

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INTRODUCTION

Pogostemon Desf. is a distinct genus in the Labiatae Juss. defined by exserted stamens and filaments usually bearing unilaterally densely arranged moniliform hairs (Bhatti, 1995). Pogostemon means 'with bearded stamen' (Stearn, 1992). Pogostemon is the largest genus in its sub-family tribe the Pogostemonoideae (Endl.) P.D. Cantino,

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Harley & Wagstaff (Cantino *et al.*, 1992) but its relationship to other included genera such as *Anisomeles* R. Br., *Colebrookia* Sm., *Comanthosphace* S. Moore, based in part on the presence of bearded stamens, has been questioned (Bhatti & Ingrouille, 1995, 1996a,b, 1997). The centre of diversity of the genus is in the Indian subcontinent and there are other lesser centres of diversity in parts of south-east Asia. The genus is of moderate size with 80–90 species currently recognized. It includes diverse species ranging from tall sub-shrubs to slender aquatic herbs. This diversity has been recognized by the description of different infra-generic classifications or by the recognition of more than one genus, but these systems have not been stable (Table 1). For example different authors at various times have recognized two segregate genera, *Dysophylla* Benth. or *Eusteralis* Raf., or only one undivided genus, *Pogostemon sensu lato*. Others have recognized similar but slightly differently delimited groups at different infra-generic ranks.

Species have been transferred between groups. In particular the boundary between Dysophylla and Pogostemon sensu stricto has been difficult to establish with different authors including different species in each group. On the basis of sharing the characters "opposite, broad and petiolate leaves, presence of crystals in the calyx and absence of an aerenchyma tissue in the stem" El-Gazzar & Watson (1967) transferred the four species, D. auricularia (L.) Blume, D. myosuroides Benth. in Wallich, D. rugosa Hook.f. and D. salicifolia Dalz. ex Hook.f. to Pogostemon. Wu & Li (1975) also transferred D. falcata (C.Y.Wu) C.Y. Wu & Li to Pogostemon for similar reasons. Press (1982) maintained a single genus Pogostemon for all species including those described in Dysophylla.

Some workers have turned to multivariate techniques to try to elucidate the patterns of relationship. Khanam *et al.* (1994) carried out a cluster analysis on 13 species of *Pogostemon* from Bangladesh, using 34 characters, 27 of them morphological and seven chemical (presence or absence of different flavonoids). They identified two main groups, one of which included six species normally placed in *Dysophylla* plus *P. strigosus* (Benth.) Benth. in DC, and the other group containing species in *Pogostemon* sect. *Paniculatae* Benth. The flavonoid characters were not useful in defining the groups.

Press (1982) included 75 species of *Pogostemon sensu lato* in his phenetic analysis of tribe *Pogostemoneae* (Benth. ex Endl.) Briq. using 64 morphological characters. In a principal coordinates analysis *Pogostemon sensu lato* was clearly separated from other taxa. There was a transition between *Pogostemon sensu stricto* and species of *Dysophylla* sect. *Verticillatae* Benth. with several intermediate species, especially from *Dysophylla* sect. *Oppositifoliae* Benth. The results from cluster analyses were broadly in agreement with those from principal coordinates analysis. Press (1982) concluded that there was "a greater degree of unity between these groups than is generally acceptable between two genera".

As the result of a detailed analysis of morphological and micromorphological variation a revised classification of the genus has been proposed (Bhatti & Ingrouille, 1997) with a more clearly defined hierarchy of infra-generic taxa. The revised classification is summarized in Table 2. It includes the recognition of a new subgenus and new sections. In this paper the cladistic and phenetic analyses of data, used in constructing the revised classification, are reported.

INFRAGENERIC RELATIONSHIPS OF POGOSTEMON

TABLE 1. History of the classification of Pogostemon sensu la	TABLE 1.	History of the	e classification	of Pogostemon sensu	lato
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labl	E 1. History of the classificat	ion of Pogostemon s	ensu lato			
	Desfontaines (1815)				
	Pogostemon described with T	ype P. plectranthoide.	5			
	Blume (182	(6)				
Dysophy	ella described with Type D. au	uricularia (=M. auri	cularia L.)			
	Bentham (18	332)				
Dysophylla	Dysophylla	Pogostemo	n Pa	gostemon		
sect. Oppositifoliae	sect. Verticillatae	sect. Panicul	latae sect.	Racemosae		
(with opposite leaves)	(with verticillate leaves) (with a pan	icle) (with	a raceme)		
	Hasskårl (18	42)				
	Pogostemon (includes	Dysophylla)				
Ra	afinesque-Schmaltz (1837)					
	Eusteralis Rafinesque-	-				
Se	thmaltz (= $Dysophylla$ sect.					
	Verticillatae Benth.)					
	Miquel (185	59)				
	Pogostemon (includes	Dysophylla)				
	Kuntze (189	91)				
	Pogostemon (including	Dysophylla)				
	Briquet (189	97)				
Dysophyl	la	P	ogostemon			
sect.	sect.	sect.	sec	sect.		
Rhabdocalicinae	Goniocalicinae	Paniculatae	Racemosae			
Calyx tube cylindrical	Calyx tube	branched	inflores	cence		
and rounded or very indistinctly pentagonal	prominently five	inflorescence				
	angled					
subsect. A. subsect. B.	subsect.	subsect.	subsect.	subsect.		
Perennial Annual	Interrupted verticils	Continuous verticils	<i>Glabriuscula</i> naked	Barbata		
species species	Verticus	verticus	filaments	hairy filaments		
Kudo	(1927)					
Dysop						
sect.	sect.					
Eudysophylla	Chotekia					
stem indumentum	stem glabrous to					
tomentose to hirsute	pubescent and leaves					
and leaves opposite,	verticillate, narrow,					
broad, ovate –	linear to linear –					
lanceolate, margin serrulate	lanceolate with an entire margin.					
scirulate						

Keng	; (1978)
Pogostemon (incl	uding Dysophylla)
Press	(1982)
sect. Verticillati (including sect. Eusteralis (Rafin.) H. Keng = Dysophylla sect. Verticillatae Benth.)	sect. Pogostemon (including Dysophylla sect. Oppositifoliae Benth.)

TABLE 1.	History	z of the	classification	of	Pogostemon	sensu	lato-	-continued

Table 2.	A summary	of a	revised	infrageneric	classification	of.	Pogostemon I)esf.

Subgenus	Section	Subsection
Pogostemon		
Allopogostemon Bhatti & Ingr.	Racemosus (Benth.) Bhatti & Ingr.	Racemosus
		Glabriusculus (Briq.) Bhatti & Ingr.
	Zygocalyx Bhatti & Ingr.	
Dysophyllus (Blume) Bhatti & Ingr.	Dysophyllus	
	Verticillatus (Benth.) Bhatti & Ingr.	

MATERIAL AND METHODS

Details of the 79 species of *Pogostemon sensu lato* analysed are listed in Appendix 1. Of the probably distinct species only *Pogostemon brevicorollus* Y.Z. Sun, *P. reticulatus* Merrill, *P. falcatus*, *P. szemacensis* (C.Y. Wu & Hsuan) Press and *P. tsiangii* (Y.Z. Sun) Press have been excluded because specimens at a comparable state of floral development were not available.

About 400 specimens were examined. Specimens are listed in Bhatti & Ingrouille (1997). Some other specimens were rejected because they were incomplete or had no information about their geographical origin. Herbarium material of *Pogostemon* has been obtained from the herbaria of the Natural History Museum (BM), the Royal Botanic Gardens, Kew (K) and the Royal Botanical Garden Edinburgh (E). More than half of the total number of species come from India. A field excursion to India was undertaken in 1991. The following herbaria of the Botanical Survey of India were visited: Poona (BSI), Calcutta (CAL), Coimbatore (MH), Dhera Dune (BSD), and Rabinath Herbarium, St. Joseph College (RHT). A few species have been observed in nature in India. These included *P. paniculatus* (Willd.) Benth. in Wallich, *P. purpurascens* Dalz. in Hook.f., *P. quadrifolius* (Benth. in Wallich) Kuntze, *P. heyneanus* Benth. in Wallich, *P. auricularius* (L.) Hassk. and *P. plectranthoides* Desf.

Species were treated as operational taxonomic units (OTUs) or terminal taxa. In 85% of species it was possible to include at least one type specimen in the analysis. Cultivated material of *P. plectranthoides* and *P. quadrifolius* was available and used to check, for the characters scored, that dried material accurately represented the living

state. In order to minimize the confusion arising from developmental plasticity, every effort was made to use mature organs for each feature. The majority of characters come from the flowers.

Both cladistic and phenetic analyses were carried out. This dual approach was chosen because at specific rank both qualitative characters used in cladistics and metric characters used in phenetics are useful. It has been recognized by many workers that cladistic analysis works best where there are a large number of characters with variable but distinct character states. Methods have been proposed to convert metric characters to discrete character states (Almeida & Bisby, 1984; Chappill, 1989; Morton *et al.*, 1995). However at species rank, in the zone between phylogenetic and tokogenetic studies, there may be relatively few morphological characters with sharply contrasting character states or characters are polymorphic. Many specific differences are expressed as differences in mean value of continuously varying characters.

The approach used here was to subject the data to the most appropriate kinds to analyses: cladistics for qualitative characters and cluster analysis and principal components analysis for mainly metric data. Where congruent results are produced, by different methods of analyses and when using different data sets, more confidence can be placed on a revised classification based upon them.

CHARACTER CODING

A mean number of five specimens was examined for each species. Only one specimen was available for 17 of the 79 species but in most species this was a type or isotype. All stems on each specimen were examined and 5–10 calyces and corollas were examined from each specimen, but, if there was any doubt about identity, or there was a large degree of variation, the number of the samples was increased. Nutlets were obtained from fruiting calyces. Details of character variation are listed in Bhatti & Ingrouille (1997).

Two data sets were prepared, one for cladistic analysis and one for phenetic analysis. 135 character states were scored and encoded as 41 multistate characters for cladistic analysis (Table 2, Fig. 1). Metric variables were screened following the methods recommended by Tabachnick & Fidell (1989). Some pairs of variables were very highly correlated. In this case, if it was possible that the variables were logically correlated so that inclusion of both variables would positively weight a feature of the plant, one of each pair of variables correlated at r > 0.85 was excluded from the phenetic analyses. Six metric variables were excluded in this way leaving 52 metric variables (Table 4). The distribution of metric characters was examined and 18 with high skewness or kurtosis were transformed to their log value to normalize them. In addition, 54 binary variables from characters not otherwise measured as a metric variable, were included by recoding multistate characters from the cladistic analysis.

Metric data can be found in Bhatti & Ingrouille (1997) and the cladistic data set is provided in Appendix 2.

CLADISTIC ANALYSIS

Character states were treated as unordered. Cladistic analysis was carried out by a combination of HENNIG86 (Farris, 1988), PAUP 3.1.1 (Swofford, 1993) and

Plant	Cl	form	herb(0)/ hydrophypte or helophyte (1)/shrub(2)
	C2	trichomes	glabrous(0)/simple-unicellular(1)/simple- multicellular(2)/branched(3)/simple and branched(4)
Leaf	C3	petiole	<pre>petiolate(0)/sub-sessile(1)/ sessile)(2)</pre>
	C4	phyllotaxis	2(0)/3(1)/4(2)/5(3)/6-10(4) leaves at each node
	C5	shape	simple(0)/compound(1)
	C6	shape	linear-linear lanceolate(0)/lanceolate(1)/elliptic to elliptic-oblong(2) /ovate(3)/orbicular(4)
	C7	apex	obtuse(0)/acute(1)/acuminate(2)
	C8	base .	rounded(0)/cuneate(1)/truncate(2)/cordate(3)
	C9	margin	dentate(0)/double-dentate(1)/crenate(2)/double- crenate(3)/serrate(4)/inciso-serrate -crenate(5)/ entire-revolute(6)
Inflorescence	C10	branching	single terminal(0)/spike/terminal spike with 2 laterals(1)/with more than 2 laterals(2)
	C11	density	lax(0)/lax below and dense $above(1)/dense(2)$
	C12	bracts	membranous and caducous(0)/not membranous
			and persistent(1)
	C13	bract shape	filiform(0)/spathulate(1)/narrow lanceolate(2)/ broad ovate(3)
	C14	bract toothing	entire(0)/entire(1)
Calyx	C15	circumference	wide(1)/equal to tube length(2)/narrow(3)
,	C16	shape	cylindrical(1)/tubular-inflated(2)/infundibular(3)
	C17	shape	symmetrical(1)/asymmetrical(2)/zygomorphic(3)
	C18	venation	5-veined(1)/5 main veins with multiple parallel
			secondaries at sinus(2)/5 main veins with a variable number of extra weaker primary veins between(3)/10-veined(4)
	C19	teeth	broadly triangular(0)/narrowly-triangular(1)/ lanceolate (2)
	C20	margin	glabrous(0)/ hairy(1)/bristly(2)
	C21	exterior	glabrous(0)/sparsely hairy(1)/densely hairy(2)
	C22	interior of teeth	glabrous(0)/tips hairy(1)/densely hairy(2)
	C23	interior tube	glabrous(0)/hairy(1)
	C24	glands	obscure(0)/obvious(1)
Corolla	C25	tube to limb	length ratio $<1(1)/=1(2)/>1(3)$
	C26	shape	length diameter ratio $\le 1.6(1)/1.7 - 1.8(2)/ \ge 1.9(3)$
	C27	tube shape	infundibular(1)/tubular(2)
	C28	upper lip length	middle lobe ≅lateral lobe length(1)/middle > lateral (2)
	C29	upper lip width	middle lobe \cong lateral lobe (1)/middle < lateral (2)
	C30	upper lip shape	lateral lobes with parallel sides(1)/ winged(2)
	C31	lip length	lower < upper(1)/ lower \cong upper(2)/ lower >
	6.00	1 1.1	upper(3) (1) (1) (1) (1) (2) (1) (1) (3) (1)
	C32	lip width	lower $\leq \frac{1}{2}$ upper(1)/ lower $\cong \frac{1}{2}$ upper(2)/ lower $> \frac{1}{2}$ upper(3)
Stamens	C33	moniliform hairs	absent(0)/ present(1)
	C34	filament bases	(glabrous(1)/villous(2)
	C35	insertion	arising at the same height in tube(1)/ at two heights(2)
Gynoecium	C36	disc	symmetrical(1)/ asymmetrical(2)
,	C37	stigma	smooth (0) / hairy(1)
	C38	nutlet surface	<pre>scales(0)/ spinulose(1)/ truminate(2)/ reticulate- foveate with secondary reticulations(3)/ reticulate- foveate(4)/ reticulate(5)/ granulate(6)/ reticulate- puncticulate(7)/ puncticulate(8)/ smooth (9)</pre>
	C39	hairs	absent(0)/ present(1)
	C40	nutlet shape	oblong with angular upper end(0)/ ovate(1)/ orbicular(2)/ oblong(3)/ obovate(4)/ lanceolate- linear-lanceolate(5)/ elliptic-elliptic-oblong(6)/
			elliptic-fusiform(7)/ D-shaped(8)

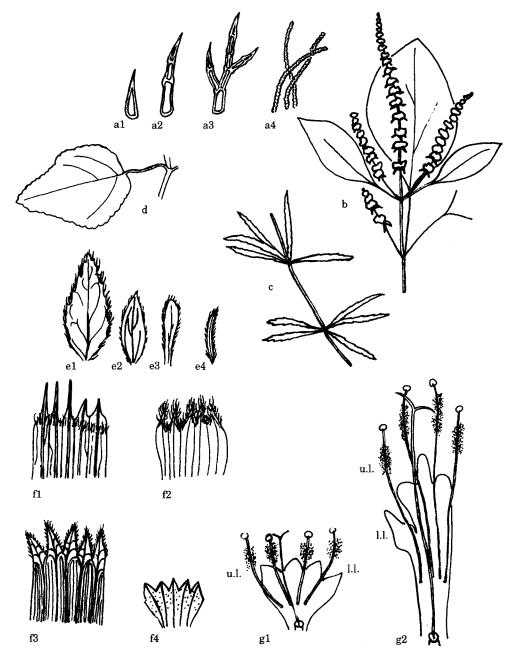


Figure 1. Qualitative characters used in cladistic and phenetic analysis: trichomes, a1 - simple, unicellular, a2 - simple, multicellular, a3 - branched, multicellular, a4 - moniliform hairs; b - inflorescence with unbranched laterals, lax throughout, leaves in pairs; <math>c - 5 leaves at each node, sessile, linear-shaped and toothed; d - petiolate, ovate with crenate margin; e - bracts, e1 - broadly lanceolate, toothed, e2 - narrowly lanceolate, even, e3 - spathulate, e4 - filiform; f - calyx (dissected), f1 - asymmetrical, tubular, ring of hairs, teeth narrowly lanceolate, 5 main veins plus others, f2 - tubular, inflated, teeth triangular margin bristly 10-veined, f3 - tubular 5 main veins and multiple parallel secondaries, f4 - infundibular, 5-veins with obvious glands; g - corolla (dissected), g1 - infundibular, upper lip (u.l) with \pm equal lateral lobes and equal to lower lip (l.l.), g2 - tubular, upper lip (u.l.) longer than lower lip (l.l.) and with lateral lobes shorter, broader than middle lobe.

P1 P2	Plant	Stem trichome length Max. no. of cells in the stem trichome
	and C1, C4 recoded	d as binary variables
P3 P4 P5 P6 P7 P8 P9 P10	Leaf	Petiole length Max. number of leaves at each node Max. length of lamina Max. length of lamina at widest point Max. width at widest point Max. width at mid point Ratio of total length and width of the lamina Ratio of total length and width of the length of lamina where lamina har widest point
P11 P12		Leaf trichome length Max. no. of cells in the leaf trichome
	and C5, C6, C7, C	8 recoded as binary variables
P13 P14 P15 P16 P17 P18 P19 P20 P21 P22	Inflorescence	Length of the terminal spike without stalk No. of verticils per terminal spike No. of the flowers on the lowermost verticil on the terminal spike Length in between first 2-verticil on the terminal spike Length in between last two verticils on the terminal spike Internode trichome length Internode trichome max. no. of cells Peduncle trichome max. no. of cells
	and C12, C13, C14	recoded as binary variables
P23 P24 P25 P26 P27 P28 P29 P30 P31 P32 P33 P34 P35	Calyx	Length of calyx in mm. Circumference Calyx circumference to length ratio Length of longest tooth Length of shortest tooth Length of the calyx tube without teeth Calyx tube and teeth length ratio Length difference in between longest and shortest tooth Max. width of tooth at the base Min. width of tooth at the base Width difference in between max. and min. width of teeth Max. length of trichome Max. no. of cells in trichome
	and C15, C16, C17	7, C18, C20, C21, C22, C23, C24 recoded as 15 binary variables
P36 P37 P38 P39 P40 P41	Corolla	Total length of the corolla Length of the lower lip of the corolla Width of the lower lip of the corolla Width of the upper lip at the sinus of the lower and upper lip Length of the central lobe of the upper lip Width of the central lobe of the upper lip
	and C27, C29, C30	0, C31, C32 recoded as binary variables
P41 P42 P43 p44 P45	Stamens	Height of the corolla tube where filaments attached Length of the longest filament Length of shortest filaments Length difference between the longest and the shortest filaments Length exserted part of the longest filament out of the corolla
	and C33, C34, C33	5 recoded as binary variables
P46 P47 P48 P49 P50 P51 P52	Gynoecium	Length of the style Length of the longest lobe of the stigma Length of the shortest lobe of the stigma Length of the disc Length of the nutlet Width of nutlet Length and width ratio
	and C36, C37, C39	9, C41 and C38 and C40, C41 recoded as binary variables

TABLE 4. Variables measured for phenetic analysis, metric characters in full, binary/ordered multistates recoded from cladistic data set (explanation in text)

McCLADE 3.04 (Maddison & Maddison, 1992). Tree searching was carried out in HENNIG86 by a combination of mhennig* followed by bb* procedures. Successive weighting was carried out using the xsteps procedure for 5 or 6 iterations. A strict consensus tree was computed by the procedure Nelsen. Tree searching on PAUP * was carried out by heuristic search using the TBR routine. Analysis was carried out to the limits of the memory of the computer. This meant, that in some analyses not all shortest trees could always be retained. Polytomous trees were retained only if a more highly resolved compatible tree did not exist. Strict and majority rule consensus trees were constructed.

PHENETIC ANALYSES

Phenetic analyses were carried out using SPSS-PC FOR WINDOWS version 6.0 (Norusis, 1985) and NT-SYS (Rohlf, 1990). Cluster analysis and principal components analysis was carried out on a data set including both metric and binary data. Characters were standardized as Z-scores to have the same range. A distance matrix of Squared Euclidean distance was calculated. This distance was used as a measure of phenetic difference.

Clustering was carried out by Ward's method of minimum variance clustering. Grouping of OTUs by Ward's method has the potential disadvantage that large clusters have a high gravity and 'attract' distinct OTUs or small groups so that they can be included in the larger cluster in a rather spurious fashion. Various other kinds of clustering of subsets of the data, for example analysing metric and binary characters separately, or using different similarity coefficients and clustering methods were also carried out but are not reported here. They produced essentially similar results to clustering by Ward's method but clusters did not correspond as well to the clades produced in the cladistic analysis.

Principal Components Analysis was also carried out because it has the advantage of abstracting factors, one of which may represent some underlying variable like overall size which may affect many characters. There were 40 significant factors but only the first four were extracted. Factor scores were calculated for each OTU and the first two plotted in a scatter diagram.

RESULTS AND DISCUSSION

Cladistic analysis of all species together revealed three major clades which were present in all shortest trees saved. The same clades were resolved using HENNIG86 with successive weighting and using PAUP 3.1.1. These clades were taxonomically recognized as the subgenera *Pogostemon*, *Allopogostemon* and *Dysophyllus*. Characters which distinguish them are listed in Table 4. Each clade was then subjected to separate analysis and rooted using the closest sister species from the other major clades/subgenera. The resulting cladograms of these separate analyses are shown

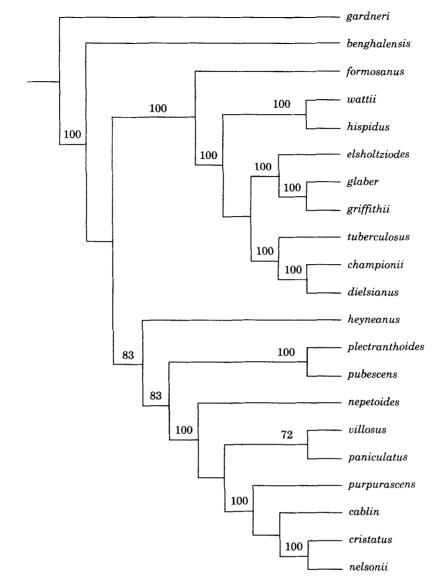


Figure 2. Cladogram of *Pogostemon* subgenus *Pogostemon* 50% majority consensus cladogram produced from 87 shortest trees rooted on *P. amaranthoides* and *P. williamsii*. Percentage of trees agreeing with branches indicated.

in Figures 2–4. Character state distributions for these cladograms can be scanned in Appendix 2 where species have been ordered in the same sequence as the cladograms. It is clear that although there are relatively few unambiguous synapomorphies, both major and minor clades have distinct suites of character states.

Analysis of major clade/subgenus *Pogostemon* (Fig. 2) produced 87 shortest trees. The 50% majority rule consensus tree divides the species into two groups but places *P. benghalensis* and *P. gardneri* as sister taxa to the rest.

Analysis of major clade/subgenus Allopogostemon produced 5200 trees. A 50% majority consensus tree is presented in Figure 3. One subclade, present in 89% of

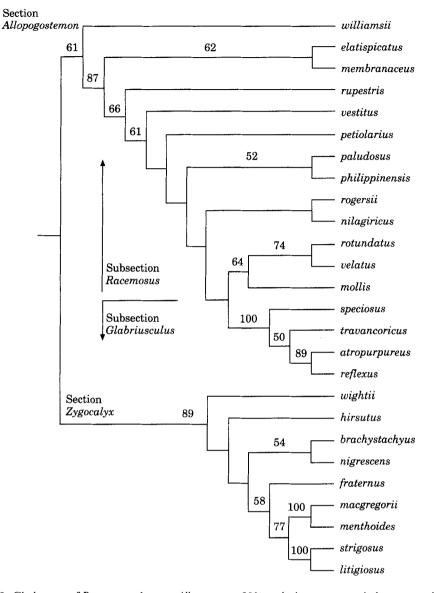


Figure 3. Cladogram of *Pogostemon* subgenus *Allopogostemon* 50% majority consensus cladogram produced from 571 shortest trees rooted on *P. amaranthoides* and *P. gardeneri*. Percentage of trees agreeing with branches indicated.

all trees, was taxonomically recognized as section Zygocalyx and another, present in 100% of all trees, as Subsection *Glabriusculus*. The remaining species were placed in a paraphyletic Subsection *Racemosus* sister to Subsection *Glabriusculus*.

Analysis of major clade/subgenus Dysophyllus (Fig. 4) produced two shortest trees which differ only in the placement of *P. crassicaulis* and *P. trinervis* either as a monophyletic clade or as a paraphyletic clade sister to a clade including *P. erectus*, *P. stellatus*, *P. koeheanus*, *P. stocksii* and *P. deccanensis*. The taxonomic treatment was to recognize one large subclade as section Verticillatus but exclude *P. tisserantii* and *P.*

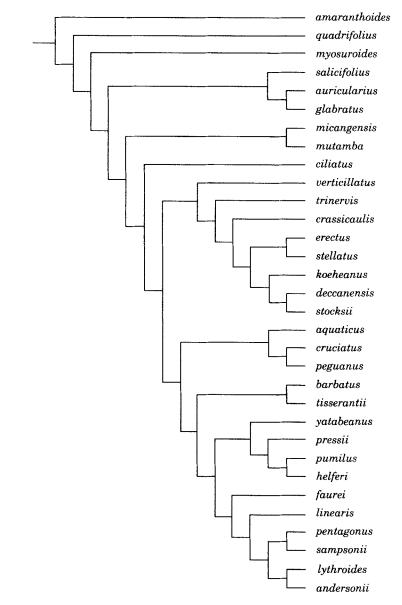


Figure 4. Cladogram of *Pogostemon* subgenus *Dysophyllus* 50% majority consensus cladogram produced one of 2 shortest trees rooted on *P. williamsii* and *P. gardeneri*.

barbatus which differ from the others in phyllotaxy or leaf shape. They were placed in section *Dysophyllus* which is here shown to be a paraphyletic group.

Results of the cluster analysis and principal components analysis correspond very closely with those from the cladistic analysis except for the placement of a few individual species. The dendrogram is shown in Figure 5. Two main clusters are produced.

The first cluster (*P. erectus-P. micangensis*) is nearly perfectly congruent with the *Pogostemon* subgenus *Dysophyllus*. Only *P. amaranthoides* is clustered elsewhere. Similarly

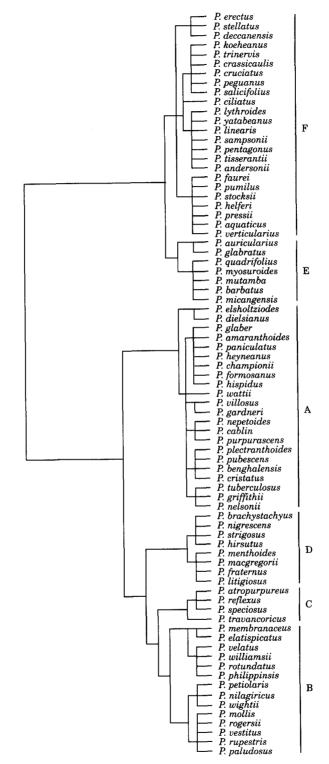


Figure 5. Phenogram of cluster analysis by Ward's method of 79 Pogostemon species. Excluding four mis-clustered species (P. salicifolius, P. barbatus, P. amaranthoides and P. wightii) clusters are A = Subgenus Pogostemon, B = Subgenus Allopogostemon Section Racemosus Subsection Racemosus, C = Subgenus Allopogostemon Section Racemosus Subsection Clabriusculus, D = Subgenus Allopogostemon Section Zygocalyx, E = Subgenus Dysophyllus Section Dysophyllus, F = Subgenus Dysophyllus Section Verticillatus.

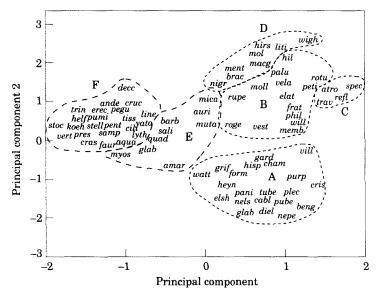


Figure 6. Scatter diagram of first (X-axis) and second (Y-axis) principal component scores (comprising 28.5% and 8.2% of the variation). OTUs coded by first letters of specific name, groups as follows; A=Subgenus Pogostemon, E=Subgenus Dysophyllus Section Dysophyllus, B=Subgenus Allopogostemon Section Racemosus Subsection Racemosus, D=Subgenus Allopogostemon Section Zygocalyx, F=Subgenus Dysophyllus Section Verticillatus, C=Subgenus Allopogostemon Section Racemosus Subsection Glabriusculus.

the subclusters *P. auricularius–P. micangensis* and *P. erectus–P. verticillatus* are highly congruent with Sections *Dysophyllus* and *Verticillatus* except *P. salicifolius* and *P. barbatus* are clustered with species of Section Verticillatus.

The second main cluster (*P. elsholtziodes–P. paludosus*) has two very distinct subclusters (*P. elsholtziodes–P. nelsonii*), labelled A, and *P. brachystachyus–P. paludosus*), labelled B, C & D, which are highly congruent with subgenera *Pogostemon* and *Allopogostemon*. The only misclustered species is *P. amaranthoides* which is included in subcluster A. Within the *P. brachystachyus–P. paludosus* subcluster smaller clusters are congruent with Subsection *Racemosus* (*P. membranaceus–P. paludosus*) labelled B, but with the inclusion of *P. wightii*, Subsection *Glabriusculus* (*P. atropurpureus–P. travancoricus*) labelled C, and Section *Zygocalyx* (*P. brachystachyus–P. litigiosus*) labelled D.

P. salicifolius, P. barbatus, P. amaranthoides and *P. wightii* are mis-clustered because in the classification emphasis was placed on the arrangement determined by cladistics since it is based on more sharply marked character states. *P. salicifolius* and *P. barbatus* clustered with species of Section Verticillatus despite having leaves in opposite pairs rather than a whorl of leaves at each node. *P. amaranthoides*, in Subgenus Dysophyllus, is misplaced in cluster A. It is a very distinct species which has floral characters like other species in Subgenus Dysophyllus but is vegetatively a large and robust plant like some species of Subgenus Pogostemon with which it clusters. For similar reasons *P. wightii* was classified in Section Zygocalyx because of its calyx shape and hairiness but is misplaced in cluster B.

The first principal component accounts for 28.5% of the variation and the second for 8.2% of the variation. The scatter diagram of the factor scores for these two components clearly shows the three major clusters/subgenera and also other groups (Fig. 6). For clarity, lines separating the taxa have been superimposed on the diagram.

Character distributions of valuable characters, including mean values and standard errors for metric variables are summarized for important characters in Table 5.

A comparison of the extent of phenetic and cladistic variation in each subgenus is interesting. Phenetic variation within and between subgenera is shown graphically in Figure 7. Differences in the mean phenetic difference within each clade are highly significant (Subgenus Dysophyllus vs. Subgenus Pogostemon $t = 5.98^{**}$ df 704, Subgenus Dysophyllus vs. Subgenus Allopogostemon $t = 11.79^{**}$ df 846, Subgenus Pogostemon vs. Subgenus Allopogostemon $t = 7.43^{**}$ df 560). Mean phenetic distances between sections are recorded in Table 6.

These results indicate that species in Subgenus Dysophyllus are not as phenetically distinct as in those in the other subgenera. A similar finding is indicated by cladistic analysis because, although the mean number of character state changes on internal branches of each of the main clades/subgenera are not significantly different, the number of changes concentrated in terminal branches of Subgenus Dysophyllus is on average 2.3 compared to 3.1 for Subgenus Pogostemon and 3.3 for Subgenus Allopogstemon. Probabilities of a significant difference are P=0.07 for the comparison Pogostemon versus Dysophyllus and P=0.03 for the comparison Allopogostemon versus Dysophyllus.

Our results are broadly in agreement with those of Press (1982) but the inclusion of several new characters, especially from the calyx, has sharpened the definition of the various groups. There is a remarkable agreement between the results of the cladistic and phenetic analyses. Most of the taxa formally recognized in the classification are monophyletic according to the cladistic analysis, but some paraphyletic groups sister to monophyletic clades were also taxonomically recognized. Character state distributions of subspecies and sections are listed in Table 5.

There is a considerable degree of homoplasy in the genus. This may be related to convergence between species which have become adapted to the same habitat. For example in Section *Racemosus*, which has eleven species in southern India, five species from the Philippines and one from southern Africa, most species are dry habitat herbs with simple inflorescences and relatively narrow bracts. The flowers are relatively large and showy for the genus, and have long filaments. The base of the filaments is normally hairy. The calyx is clearly marked by 10 parallel veins.

Subsection *Glabriusculus* is unusual because moniliform hairs are absent from the filaments, though a few simple hairs can sometimes be found, and also the ovary/nutlets have stalked glands. *P. paludosus* Benth. in Wall., from the sister Subsection *Racemosus*, also has glands on its nutlets, but has moniliform hairs. This may indicate either parallel evolution of nutlet glands or that their presence is ancestral but they have been lost from most other members of their section.

In Section Zygocalyx the species have a two-lipped calyx, a condition which is seen at its most extreme in *P. litigiosus*. The teeth are often rather subulate and the interior of the calyx is fringed by a ring of hairs, which is either continuous or broken and is sometimes expressed as tufts of hairs at the sinus of each pair of teeth or inside the teeth. This section is centred geographically on the region of eastern Himalayas to Yunnan and Thailand but includes three endemics to Java, Borneo and Sri Lanka respectively. Parallel evolution is observed in *P. reflexus* and *P. speciosus* of Subsection *Glabriusculus*, which have a calyx with some teeth rather subulate and fringed by bristles rather like that in Section Zygocalyx. Alternatively, *P. menthoides* of Section *Zygocalyx* has relatively few moniliform hairs rather like species of Subsection *Glabriusculus*.

	subg. Allopogostemon	g. stemon	subg. Pogastenion	su Dyse	subg. Dysophyllus
Character	sect. <i>Racemosus</i>	sect. Zygocalyx		sect. Dysophyllus	sect. Verticillatus
number of species surveyed	17	6	21	10	22
Habit	herb	ţ	shrubby	herb	herb
Phyllotaxis	2 leaves at each node	each node	2 leaves at each node	2-4 leaves at each node	>2 at cach node
Petiole	petiolate	late	petiolate	sessile- petiolate	scssile
Leaf length/ width ratio	2.34 (0.14)	2.29 (0.20)	2.48 (0.24)	4.27 (0.80)	9.28 (1.42)
Leaf margin	crenate or scrrate	r scirate	double dentate	dentate or serrate	dentate
Inflorescence	unbranched verticillaster	verticillaster	branched verticillaster	unbranched verticillaster	unbranched verticillaster
Calyx shape	tubular teeth triangular	± two-lipped teeth awl shaped	tubular inflated	usually campanulate	campanulate
Calyx length	5.0 (0.3)	4.7 (0.3)	4.3 (0.2)	2.3 (0.2)	1.8 (0.1)
Calyx throat circumf./tube length ratio	0.89 (0.04)	0.88 (0.05)	0.86 (0.03)	1.32 (0.04)	1.46 (0.05)

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continued

TABLE 9: CHIMARCE STATES AND THEADS OF LADGES HILLA "GENELIA" (ANA. MATHARI ETFOIS III DATETHILESIS - COMMUNIC	s 5 main veins 5 veins te and multiple intercostal region parallel translucent veins	d glabrous or ±glabrous hairy but hairs not in a ring	6.4 3.6 (0.4) (0.5)	6.3 3.1 (0.4) (0.4)	2.7 2.3 (0.10) (0.10)	1.7 1.8 (0.09) (0.15)	1.2 1.0 (0.02) (0.01)	11.8 4.4 (0.7) (0.3)
THE STATES AND THEATS OF CLAUCE	veins 5 main veins aarallel plus reticulate al veins intercostal veins	or with throat ringed of hairs by hairs top of d teeth	5.6 0.7) 5.6 (0.4)	3 5.4 4) (0.5)	3.2 3.0 (0.05) (0.07)	2.3 1.9 (0.18) . (0.17)	1.2 1.2 0.02) (0.04)	2.3 11.4 1.0) (1.5)
LABLE J. CIIIAI A	Number of 5 main veins calyx veins plus 5 parallel intercostal veins	Cabyx interior glabrous or with indumentum a ring of hairs around top of tube and teeth	Filament 8. length mm (0.	Corolla length 7.3 (0.4) mm	Corolla lower 3. lobe length (0. mm	Corolla upper 2. lobe width (0. mm	Corolla lower 1. lip length/width (0.	Style length 12.3 mm (1.0)

INFRAGENERIC RELATIONSHIPS OF POGOSTEMON

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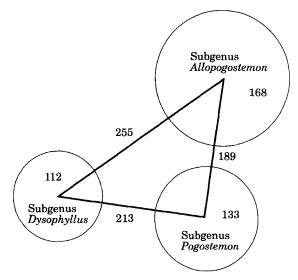


Figure 7. Cluster diagram showing mean phenetic distance (squared euclidean distance) within and between the three subgenera.

	Subgenus A	llopogostemon	Subgenus Pogostemon	Subgenus Dysophyllus		
	Section Racemosus	Section Zygocalyx		Section Dysophyllus	Section Verticillatus	
Number of species	17	9	21	10	22	
mean phenetic distance within and between sections (SE)						
Section Racemosus	154 (4.5)					
Section Zygocalyx	191 (4.0)	125 (7.4)				
Subgenus Pogostemon	189 (2.9)	188 (2.8)	133 (3.2)			
Section Dysophyllus	222 (5.7)	192 (5.2)	183 (3.2)	121 (3.2)		
Section Verticillatus	269 (4.1)	233 (4.2)	226 (2.5)	133 (2.4)	91(2.5)	

TABLE 6. Variation within and between groups. Phenetic distance calculated from squared euclidean distance between all pairs of species

Subgenus Pogostemon includes most of the shrubby species of Pogostemon including the type species of the genus P. plectranthoides. Many are of economic importance as the source of Patchouli, the pungent oil extracted from dried plants. The shrubby Patchouli species include several rather poorly defined taxa around P. benghalensis (Burm.f.) Kuntze, P. plectranthoides, P. cablin (Blanco) Benth. and P. heyneanus. Patterns of variation are complicated by the cultivation of variants which rarely flower and are maintained vegetatively, and by the introduction of cultivars into areas outside their native distribution. New data from a range of populations of each species are required to ascertain whether currently recognized species should be considered as infra-specific variants of one or more polymorphic species. Downloaded from https://academic.oup.com/botlinnean/article/128/2/159/2557274 by guest on 20 April 2024

Similarly, a population based study is needed to evaluate the relationships of species in Subgenus Dysophyllus, many of which are adapted as aquatic or semi-aquatic herbs. The taxonomic status of geographically narrowly restricted endemics like *P. erectus* (Kuntze) Press from Bombay, *P. koehneanus* (Muschler) Press from Thailand and *P. andersonii* (Prain) Press from Sikkim, to geographically widespread taxa like *P. stellatus* (Benth.) Bhatti and Ing., *P. crassicaulis* (Benth. in Wallich) Press and *P. cruciatus* (Benth. in Wallich) Kuntze would be especially valuable.

Placing the boundary between Subgenus Dysophyllus and the rest of the genus has taxed previous workers but here emphasis has been placed on calyx characters so that, for example, *P. amaranthoides* is placed within the subgenus as sister to the rest of the subgenus, because it too has a broadly conical and translucent calyx. Nevertheless a taxonomic problem remains because a number of very distinct taxa have been lumped together rather artificially in Section Dysophyllus which is paraphyletic to Section Verticillatus. These include the type species of the subgenus, *P. auricularius*. Different species of Section Dysophyllus have different combinations of character states found in Section Verticillatus and in the other subgenera. *P. mutamba* and *P. micangensis*, the African species, have normally been placed in Pogostemon species but although they also have a calyx with intercostal veins it is very strongly conical/ campanulate like other species of Subgenus Dysophyllus. Another problematical African species is *P. tisserantii* from Chad. It is unique in Pogostemon in having a pinnatifid leaf.

Most species of Subgenus *Dysophyllus* have a small corolla in which the separate lobes of the upper lip and the single lobe of the lower lip are approximately equal in size and shape so that the corolla has a symmetry which is nearly actinomorphic. The evolution of this kind of corolla is also found as a parallelism in other genera of Labiatae which have many aquatic or marsh species, like *Mentha* L. Many species in Subgenus *Dysophyllus* have weak stems and most have several linear to linearlanceolate leaves arranged in a whorl at each node.

Part of the taxonomic difficulty experienced in placing the boundary between infra-generic groups in *Pogostemon* is because of the lack of a reasonable outgroup to establish where the root of the cladogram is and to establish ancestral states. The absence of a good outgroup makes it difficult to translate directly the cladogram into a phylogeny for *Pogostemon*.

Other genera of subfamily *Pogostemonoideae* do not have moniliform hairs on the filaments and are unlikely to be phylogenetically related (Bhatti & Ingrouille, 1995). A proper outgroup for *Pogostemon* may not be found in the Labiatae but in the Verbenaceae Jaume St-Hil. where very similar moniliform hairs have been recorded, not on the filaments, but on other floral parts (Endress, 1994). If the shrubby Verbenaceae contain the most recent ancestors of *Pogostemon* it seems likely that it is the subshrubs of subg. *Pogostemon* which are sister to the other clades in the genus. The other subgenera include representives with various specializations, many involving reductions or loss. For example the aquatic and marshland herbs of Subgenus *Dysophyllus* have weak stems, narrow leaves and small campanulate flowers. It is also possible that evolutionary convergence, especially in marshland and aquatic species, has given rise to group of heterogeneous taxa which share some characters. The dry habitat herb species of Subgenus *Allopogostemon* have pollination and nutlet dispersal adaptations such as the loss of moniliform hairs, a zygomorphic corolla with subulate teeth and the evolution of glands on the nutlets.

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APPENDIX 1

A revised classification of Pogostemon (Bhatti & Ingrouille, 1997) with species studied and including synonymy and geographical origin.

Pogostemon Desf.

Subgenus Pogostemon

P. atropurpureus Benth. in A. DC. Prodromus 12: 154. Paris (1848). (P. imberbe Wright, ex Hook. f. Fl. Brit. Ind. iv. 637 (1885). India.

P. benghalensis (Burm.f.) Kuntze, Revisio generum plantarum 2: 529 (1891). (Origanum benghalense Burm. f. Fl. Ind. 128: 38 (1768), Pogostemon paroiflorus Benth. in Wall., 1: 31 (1830), P. intermedius Benth. in Wall. Cat. n.2327 (1830), P. frutescens Graham Cat. Bomb. Pl. 149, (1839) pro parte, P. purpuricaulis Dalz. pro parte in Hook. Kew Journal 2: 337 (1850), P. plectranthoides auct. pro maj., non Desf.). Nepal, India, Bangladesh, Burma, Thailand and Sri Lanka.

P. cablin (Blanco) Benth. in A. DC., Prodromus 12: 146. Paris (1848). (Mentha cablin Blanco Fl. Philip. 473 (1837), Mentha auricularia Blanco, op. cit. ed. 2 (1845) non Linn., P. patchouly Pellet. in Mém. Soc. Sci. Orléans, v. 277.t.7 (1845) pro parte, P. suavis Ten. in Parl. Giornale Botanico Italiano. Firenze. ii, 56 (1847) pro parte). Widespread in the Asian Tropics to Fiji. Introduced into many places as one of the sources of Patchouli oil.

P. championii Prain in Kew Bull. 254 (1908). Hong Kong.

P. cristatus Hassk. in Hoev. & De Vriese, Tijdschr. 10: 127 (1843). Timor, Lesser Sunda Islands.

P. dielsianus Dunn in Notes of the Royal Botanic Gardens Edinburgh 8: 159 (1913). Western China.

P. elsholtzioides Benth. in A. de Candolle, Prodromus 12:153. Paris (1848). The Himalayas.

P. formosanus Oliver in Hook.f. Icones 25, Pl. 2440 (1896). Taiwan.

P. gardneri Hook.f. in Flora of British India 4: 632. (1885). Western India.

P. glaber Benth. in Wall. Plantae Asiaticae Rariores 1:31.(1830). North-western India and Nepal to Yunnan and Thailand.

P. griffithii Prain in Kew. Bull. 181, (1908). Burma.

P. heyneanus Benth. in Wall. Plantae Asiaticae Rariores 1: 30. (1830). (P. patchouly Pellet. in Mém. Soc. Sci. Orléans, 277: 7 (1845) pro parte, P. suavis Tenore in Parl. Giornale Botanico Italiano. Firenze. ii, 56 (1847) pro parte). Widespread in southern Asia from Sri Lanka to Indonesia. Introduced into many areas including the Seychelles as a source of Patchouli oil.

P. hispidus Prain in Kew Bull. 254, (1908). Thailand to Bangladesh.

P. nelsonii Doan in Humbert, Fl. Gen. Indo-Chine 4: 975 (1936). Southern India, Kerala.

P. nepetoides Stapf. in Kew Bull. 116 (1908). The Philippines, Luzon.

P. paniculatus (Willd.) Benth. in Wall. Plantae Asiaticae Rariores 1: 30. (1830). (Elsholtzia paniculata Willd. Sp. Pl. 3: 59. (1800)). Southern India and Thailand.

P. plectranthoides Desí. in Mem. Mus. Natl. Hist. nat. Paris 2: 154.t.6 (1815). (Mentha secunda Roxb. Hortus Bengalensis 44. Serampore (1814)). India, Bangladesh.

P. pubescens Benth. in A. DC., Prodromus 12: 152. Paris (1848). South-east Asia.

P. purpurascens Dalz. in Hooker's 7. Bot. 2: 337 (1850). India from Bombay to Assam.

P. tuberculosus Benth. in Wall. Plantae Asiaticae Rariores 1: 31 (1830). Eastern Himalayas.

P. villosus Benth. in Wall., Plantae Asiaticae Rariores 1: 30 (1830). India, Bengal and Bangladesh.

P. wattii C.B. Clarke, J. Linn. Soc. 25: 59. (1889). (P. battakianus Ridl. in Journal of the Asiatic Society. Calcutta. 1: 85). India, in western Assam near Burma.

Subgenus Allopogostemon Bhatti & Ingr. Bulletin of the Natural History Museum. London. (Botany) 27(2): 97 (1997). (Pogostemon section Racemosa Benth. pro parte).

Section Racemosus (Benth.) Bhatti & Ingr. Bulletin of the Natural History Museum. London. (Botany) 27(2): 97 (1997). (Pogostemon section Racemosa Benth., Pogostemon section Barbata Briq. pro parte).

Subsection Racemosus

P. elatispicatus Bhatti & Ingr. Bulletin of the Natural History Museum. London. (Botany) 27(2): 102 (1997). Philippines.

P. membranaceus Merr. in Philippin Journal of Science (Botany) 7: 347 (1912). Philippines.

P. mollis Benth. in Labiatarum genera et species, 155 (1832-6) London. Southern India in the western Ghats.

P. nilagiricus Gamble, Fl. Madras. 1134 (1924). Southern India, Nilagiri (Kunda Hills near Ootacamund).

P. paludosus Benth. in A. DC., Prodromus 12: 154. Paris (1848). India.

P. petiolaris Benth. in A. DC., Prodromus 12: 154. Paris (1848). Southern India in the western Ghats.

P. philippinensis S. Moore, J. Bot. 43 (1905). East Indies, Philippines and Marianas Islands.

P. rogersii N.E. Brown in Kete Bull. 379 (1909). Southern Africa.

P. rotundatus Benth. in Wall., Plantae Asiaticae Rariores 1: 31 (1830). Southern India.

P. rupestris Benth. Labiatae genera et species, 156 (1832–6) London. (non Dysophylla rupestris Dalz. in Hook. Kew Journal 3, 120 (1851)). Sri Lanka.

P. velatus Benth. in A. DC., Prodromus 12: 155. Paris (1848). Philippines.

P. vestitus Benth. in Wall. Plantae Asiaticae Rariores 1: 31 (1830). Southern India, western Tamil Nadu. P. williamsii Elmer, Leaft. Philipp. Bot. ix. 3197 (1934). Phillippines.

Subsection Glabriusculus (Briq.) Bhatti & Ingrouille Bulletin of the Natural History Museum. London. (Botany) 27(2): 102 (1997).

P. atropurpureus Benth. in A. DC., Prodr. 12: 154 (1848) (P. imberbe C.H. Wright ex Hook. f. Flora of British India 4: 637).

P. reflexus Benth. in A. DC., Prodromus 12: 155. Paris (1848). Sri Lanka.

P. speciosus Benth. in Wall., Plantae Asiaticae Rariores, 1: 31 (1830). India, Nilaghiri (Kunda Hills) on the border of Kerala and Tamil Nadu.

P. travancoricus Bedd., Icones Plantarum Indiae Orientalis 1: 34 (1869-1874). Southern India.

Section Zygocalyx Bhatti & Ingr. Bulletin of the Natural History Museum. London. (Botany) 27(2): 104 (1997).

P. brachystachyus Benth. in A. DC., Prodromus 12: 156. Paris (1848). Northern India to Burma.

P. fraternus Miq. Florae Indiae Batavae 2: 635 (1859). Northern India to China and Thailand.

P. hirsutus Benth. in Labiatarum genera et species, London. 155 (1832-6). Sri Lanka.

P. litigiosus Doan in Humbert, Flora General of Indo-China. 4: 972 (1936). Malaysia, Mt. Kinabalu only.

P. macgregorii W.W. Smith, in Record of the Botanical Survey of India. Calcutta. 6: 39 (1914). Northern Thailand.

P. menthoides Blume, Bijdragen tot de Flora van Nederlandsch Indië 3: 825 (1826). India and Burma to Indonesia and Vietnam.

P. nigrescens Dunn in Notes of the Royal Botanic Gardens Edinburgh 37: 159 (1918). Western China.

P. strigosus (Benth.) Benth. in A. DC., Prodromus 12: 155. Paris (1848). Assam, India and north-eastern Bangladesh.

P. wightii Benth. in Labiatarum genera et species, London. 156 (1832-6). Southern India.

Subgenus Dysophyllus (Blume) Bhatti & Ingr. (Dysophylla Blume (1826) pro parte).

Section Dysophyllus

P. amaranthoides Benth. in A. DC., Prodromus 12: 153. Paris (1848). Eastern Himalayas.

P. auricularius (L.) Hassk. Tijdschrift voor Natuurlijke Geschiedenis en Physiologie. Amsterdam 10: 127 (1843). (Mentha auricularia L. Mant. pl. 81 (1767); Dysophylla auricularia (L.) Blume Bijdr.: 825 (1826)). Southern Asia from Sri Lanka to eastern China and New Guinea.

P. barbatus Bhatti & Ingr., Bulletin of the Natural History Museum. London. (Botany) 27: 108 Western China to Hong Kong.

P. glabratus Chermsir. ex Press in Bulletin of the Natural History Museum. London. (Botany) 10: 73-74 (1982). Thailand.

P. micangensis G.Taylor in Journal of Botany. London. 69: Supp. 2, 166 (1931). Angola, Cameroon.

P. mutamba (Heirn) G. Taylor in 7. Bot. Lond. 69, supp. 2: 166 (1930). South-western Africa.

P. myosuroides (Benth.) Kuntze, Revisio generum plantarum 2: 530 (1891). Eastern India.

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APPENDIX 2

Data encoded for cladistic analysis. Columns ordered in sequence C1-C41. Species ordered in the same sequence as Figs 2-4.

same sequence as rigs 2-4.									
P. gardneri	02000	32011	23012	12023	22113	22222	21111	00806	0
P. benghalensis	22000	32012	13121	12022	22113	32222	21112	00804	0
P. formosanus	22000	32012	22022	12022	21112	22222	21112	00404	0
P. wattii	22000	31212	22012	12022	22113	11222	21111	00401	0
P. hispidus	22000	11041	32022	12022	22113	12221	21111	00402	0
P. elsholtzioides	22000	22001	32022	12022	21112	21222	21111	00802	0
P. griffithii	22020	01001	32022	12023	21113	22222	22111	00203	0
P. tuberculosus	23000	31032	12021	12023	11112	21222	21111	00603	0
P. championii	22000	32031	12031	12023	21113	22221	22112	00402	0
P. dielsianus	22000	32022	12031	12023	21123	32221	21112	00205	0
P. heyneanus	22000	32042	23032	12023	22113	22222	11120	00802	0
P. plectranthoides	22020	01142	23022	12022	21112	21221	21111	00804	0
P. pubescens	22000	01142	23022	12022	22112	22222	21111	00804	0
P. nepetoides	22000	30222	23122	12022	22113	22222	21111	00806	0
P. villosus	22000	31022	23122	12023	22113	21222	22111	00803	0
P. paniculatus	22000	01062	33122	12023	22113	22221	22111	00806	0
P. purpurascens	22000	11040	33122	12022	22112	32222	22111	00706	0
P. cablin	22000	30012	23132	12023	22111	32222	22111	00406	0
P. cristatus	12000	31051	23022	12022	22113	32221	22111	00403	0
P. nelsonii	00000	31052	22122	12022	11123	32222	21111	00403	0
F S (1)()									
P. williamsii	04000	32020	20022	14022	11113	21222	21122	00106	0
P. elatispicatus	04000	12040	20021	14023	11113	22222	21122	00408	0
P. membranaceus	02000	31000	20031	14023	31113	32112	11122	00508	0
P. rupestris	12000	01200	30021	14022	11113	11221	21122	00402	0
P. vestitus	02000	31100	10021	14012	21113	31222	22122	00403	0
P. petiolarius	02000	31200	10021	14022	31113	22222	22122	00302	0
P. paludosus	02000	30210	20011	14022	11113	22222	22122	00502	1
P. philippinensis	02000	31110	20021	14022	21113	22221	22122	00503	0
P. rogersii	02000	41032	10021	14022	11111	22221	22122	00402	0
P. nilagiricus	02000	31130	10021	14023	11112	22221	22122	00302	0
P. rotundatus	02000	30100	10021	14023	11113	22222	21122	00003	0
P. velatus	04100	30100	10021	14023	11113	21222	21122	00103	0
P. mollis	02000	20100	30022	14123	11113	22222	22122	00402	0
P. speciosus	02000	00160	20031	14023	31112	23222	21022	00501	1
P. travancoricus	02000	32020	10021	14013	11113	23221	21022	00503	1
P. atropurpureus	12000	21130	10021	14023	31112	23221	21021	00503	1
P. reflexus	02000	21240	10021	14023	31113	23221	21021	00503	1
P. wightii	02000	31230	20022	33132	11213	13222	21122	00302	0
P. hirsutus	02000	31010	20022	33133	11213	22222	11220	00502	0
P. brachystachyus	02000	32000	20022	33133	11211	22221	22111	00502	0
P. nigrescens	02100	30021	10022	33133	11213	22222	21111	00504	0
P. fraternus	02000	31052	20032	33132	11211	32222	22112	00402	0
P. macgregorii	02000	31120	10022	33132	11221	22221	21121	00402	0
P. menthoides	02000	31040	30022	33132	11222	22222	21121	00402	0
P. strigosus	02120	01260	30032	33132	11213	22221	21111	00104	0
P. litigiosus	12020	01260	30022	33133	11211	22222	22112	00402	0
P. amaranthoides	02000	32011	22012	11022	11123	11221	21111	00803	0
P. quadrifolius	02120	32030	12013	11023	11123	11221	21111	00103	0
P. myosuroides	02100	31011	22013	11023	11123	11111	11111	00106	0
P. salicifolius	02100	01040	32023	11023	11123	11121	11111	00903	0

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				communa					
P. auricularius	02100	11040	32013	11021	11123	11111	11111	00507	0
P. glabratus	02100	11000	22013	11021	11122	11112	11111	00407	0
P. micangensis	12100	31010	20013	21023	11123	22111	12112	00406	0
P. mutamba	02100	01102	30013	21023	11123	22111	22112	00403	0
P. ciliatus	02220	01060	30013	11021	11123	11111	22111	01613	0
P. verticillatus	12220	01202	31013	11023	11122	11111	12111	00606	0
P. trinervis	12210	40100	31013	11023	11122	11112	11111	00903	0
P. crassicaulis	12210	01001	31013	11023	11121	11111	11111	00903	0
P. erectus	12240	01200	31013	11023	21122	11111	11111	10903	0
P. stellatus	12240	02260	31012	11023	11122	11111	11111	10903	0
P. koeheanus	11210	01000	31013	11023	11122	11221	21111	00903	0
P. deccanensis	12240	11210	11013	11023	11211	12212	11111	00502	0
P. stocksii	12240	12240	20013	21023	11122	11221	21111	00903	0
P. aquaticus	12220	01062	31013	11022	11122	11111	11111	00903	0
P. cruciatus	12210	00060	30013	11022	11122	11111	11111	00903	0
P. peguanus	11220	00260	30013	11022	11112	11111	11111	00903	0
P. barbatus	12000	21020	32013	11021	11122	11111	11111	00403	0
P. tisserantii	12021	21060	32013	21023	11122	11111	11111	00402	0
P. yatabeanus	12210	01240	21012	11023	11123	11111	21111	00403	0
P. pressii	12220	31110	21012	11013	11122	11111	11111	00903	0
P. pumilus	12220	31210	21012	21023	11123	11111	11111	00703	0
P. helferi	12220	31210	21013	21022	11123	11111	11111	00903	0
P. faurei	12220	31050	21013	11022	11121	11111	11111	00703	0
P. linearis	12220	31010	20013	11023	11121	11111	11111	00406	0
P. pentagonus	12220	30210	10013	11023	11121	21111	11111	00404	0
P. sampsonii	12210	30230	11013	11023	11122	11111	11111	00403	0
P. lythroides	12220	31240	20013	10232	11221	11111	11110	00403	0
P. andersonii	12200	11200	30013	11023	21121	11111	11111	00403	0

APPENDIX 2-continued